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㉒ **An enzymatic detergent additive, a detergent, and a washing method.**

㉓ Enzymatic detergent additive the active component of which is a microbially produced lipase, wherein the lipase is producible by means of a lipase producing strain of *Pseudomonas cepacia*.

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"An Enzymatic Detergent Additive, a Detergent, and a Washing Method"

The field comprising enzymatic additives in detergents has been rapidly growing during the last decades. Reference is made to e.g. the article "How Enzymes Got into Detergents", vol. 12, Developments in Industrial Microbiology, a publication of the Society for Industrial Microbiology, American Institute of Biological Sciences, Washington, D.C. 1971, by Claus Dambmann, Poul Holm, Villy Jensen and Mogens Hilmer Nielsen.

The most common enzymatic detergent additive is a proteolytic additive, but also lipolytic detergent additives are described, e.g. in US patent No. 4,011,169, column 4, line 65 to column 5, line 68, and British patent No. 1,293,613, page 2, lines 6 to 20.

Also, a comprehensive review article of lipases as detergent additives written by Hans Andree et al. is to be found in the Journal of Applied Biochemistry, 2, 218-229 (1980), entitled "Lipases as Detergent Components".

Among the known lipases used as detergent additives to the best of our knowledge the *Fusarium oxysporum* lipase has the best lipolytic characteristics, looked upon from a detergent application point of view, vide our European patent application with publication No. 0 130 064, especially the comparative table on page 27.

If the washing process is conducted at high temperature and high alkalinity, the majority of the fat containing dirt will be removed anyway. However, mainly due to the energy crisis and the fact that many modern-day fabrics have to be washed at low temperature, low temperature washing processes (around 60°C and below) are generally preferred, and at these low temperatures the known lipases are able to dissolve only a part of the fat containing dirt.

Hitherto the efficiency of lipolytic detergent additives usually has been measured by means of EMPA (Eidgenössische Materialprüfungs-und Versuchsanstalt, St. Gallen, Switzerland) swatches Nos. 101 (olive oil/cotton) and 102 (olive oil/wood) by adaptation of the procedure described in British patent No. 1,361,386, (especially pages 4 and 7) and US patent No. 3,723,250 (especially col. 15-19). In this way lipolytic cleaning efficiency can be expressed as the differential remittance value ΔR . However, it has been found that a more relevant measure for the lipolytic cleaning efficiency or the lipolytic detergency action is the determination of the amount of residual fat and residual fat decomposition products in the laundry after the lipolytic treatment comprising a lipolytic wash as compared to the amount of fat in the laundry before the lipolytic treatment, because this is a

more direct and relevant effect of the lipolytic detergency action than the differential remittance value ΔR . By lipolytic detergency action we understand the detergency action caused by lipolytic activity. By use of this determination it has been found that even the best known detergent lipase exhibits a lipolytic detergency effect which is open to improvement.

Thus, a need exists for a lipolytic detergent additive which exhibits a considerably better lipolytic detergency efficiency at economically reasonable lipase activities in the washing solution.

Now, according to the first aspect of the invention, a lipolytic detergent additive has been found which exhibits a considerably better lipolytic detergency efficiency at economically reasonable lipase activities in the washing solution, this lipolytic detergent additive being characterized by the fact that the lipase is producible by means of a lipase producing strain of *Pseudomonas cepacia*.

It is described in Japanese patent application No. 135999 from 1980 that *Pseudomonas cepacia* is a lipase producer. However, it does not appear from this Japanese patent application that the *Pseudomonas cepacia* lipase, as described in Japanese patent application No. 135999 from 1980, is suited as an active component in an enzymatic detergent additive.

Also, in US patent No. 3,950,277 it is described in general terms that lipases from the genus *Pseudomonas* are suited as agents for removal of oily stains from fabrics, if used together with a special group of lipase activators. Also, it appears from this US patent that the lipases in consideration are well suited for pre-soaking of fabrics exclusively, whereas the enzymatic detergent additive according to the invention is well suited both as an ingredient in a presoaking detergent and in a main wash detergent.

Also, it is described in UK patent No. 1372034 that the lipase produced by means of *Pseudomonas stutzeri* ATCC 19154 is well suited as an enzymatic detergent additive in a detergent comprising specified further detergent constituents. It has been found, however, that the lipolytic detergency action of the enzymatic detergent additive according to the invention is superior to the lipolytic detergency action of the enzymatic detergent additive described in the UK patent.

It is to be understood that lipases which are immunologically identical to the lipases defined above are covered by the invention.

Also, it is to be understood that lipases deviating from the lipases defined above, but produced by genetic engineering on the basis of *Pseudomonas cepacia*, and with active centers identical to the active centers of the lipases producible by means of a lipase producing strain of *Pseudomonas cepacia*, are also covered by the invention.

Some strains belonging to *Pseudomonas cepacia* are poor lipase producers. For the purposes of this invention, a lipase producing strain of *Pseudomonas cepacia* is defined as a strain which produces more than 10 LU/ml or more than 2 NLU_{8.5}/ml (the LU and NLU_{8.5} being the lipase units defined later in this specification) in a suitable medium, e.g. a medium designated LIP7.

The medium LIP7 intended for shaking flasks is prepared with the following ingredients in grams per litre:

PHARMAMEDIA 20

corn steep liquor 40

glucose 10

soybean oil 40

pH adjusted to 7.4 before autoclaving

final pH 7.0

The composition of PHARMAMEDIA is described in Traders' Guide to Fermentation Media Formulation, 1980, Traders Oil Mill Co., p. 50 -51.

Sterilization took place at around 130°C for around 40 minutes. A 500 ml Erlenmeyer flask with 100 ml of substrate was inoculated with cells from an agar slant previously inoculated with the strain to be tested for lipase production. The flasks were shaken at 200 -250 rpm and at around 26°C for 4 days, whereafter the lipase yield was determined.

Thus, it has surprisingly been found that the enzymatic detergent additive according to the invention exhibits an improved lipolytic cleaning efficiency which will appear from documentation presented later in this specification.

In this specification with claims two (or three) different lipase activity units are used, i.e. the Lipase Unit (LU) described in AF 95/5-GB, and the NOVO Lipase Unit (NLU) described in AF 182/3-GB, both AF documents being available on request from NOVO INDUSTRI A/S, Novo Alle, 2880 Bagsvaerd, Denmark, and furthermore a modified Novo Lipase Unit, differing only from the Novo Lipase Unit (determined at pH 7.0 or closely to pH 7.0) by the fact that the activity determination is carried out at pH 8.5, this modified Novo Lipase Unit for the sake of brevity being designated NLU_{8.5}. The LU

method is a quick and simple activity determination method based on a substrate fat with a short chain, i.e. tributyrin, and thus not representative for soiling fats on textiles; the NLU method is a more elaborate activity determination method based on olive oil, which is a mixture of triglycerides containing longer chain fatty acids, considered more representative for soiling fats on textiles.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention the strain is *Pseudomonas cepacia* DSM 3333 to 3337 inclusive, or 3401, preferably 3335 or 3401. It has been found that these strains give rise to a relatively high yield of lipase.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention, the additive is provided as a non-dusting granulate. Granulates can be produced in several different ways. Reference can be made to GB patent No. 1,362,365 which describes the production of enzyme containing granulates used as detergent additives by means of an apparatus comprising an extruder and a spheronizer (sold as MARUMERIZER®), and to US patent No. 4,106,991 which describes the production of enzyme containing granulates used as detergent additives by means of a drum granulator.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention the additive is provided as a liquid. Liquid detergents exhibits a growing popularity due to the ease of application.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention, the additive is provided as a liquid with an enzyme stabilizer. The stabilizer can be propylene glycol or other agents known as stabilizers for enzyme solutions. Liquid detergents exhibit a growing popularity due to the ease of application.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention, the lipase activity is above about 2500 NLU/g of additive. In this manner, a convenient lipase activity is generated in the washing solution when the detergent additive is added to the detergent in an amount of 0.01 to 50.0 g/100 g of detergent, and when the detergent is added to the washing solution in an amount of 1 -20 g of detergent/l of washing solution.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention, the additive contains a proteolytic enzyme besides the lipase. As such proteolytic enzyme the proteolytic enzyme ALCALASE® from NOVO INDUSTRI A/S, manufactured microbially by means of *Bacillus licheniformis*, can be used. The mixed enzymatic additive can be prepared either by mixing a previously prepared granulate of proteinase

with a previously prepared granulate of lipase, or by mixing a concentrate of proteinase with a concentrate of lipase and then introducing this mixture into a granulating device, together with the usual granulating aids.

In a specially preferred embodiment of the enzymatic detergent additive according to the invention, the proteolytic activity is between about 0.5 and about 3.0 Anson Units/g of additive. In this manner, a convenient proteolytic activity is generated in the washing solution when the detergent additive is added to the detergent in an amount of 0.2 -2 g/100 g of detergent, and when the detergent is added to the washing solution in an amount of 1 -20 g of detergent/l of washing solution.

The second aspect of the invention comprises a detergent comprising an enzymatic detergent additive, the active component of which is a microbially produced lipase, wherein the enzymatic detergent additive is the enzymatic detergent additive according to the invention.

In a specially preferred embodiment of the detergent according to the invention, the detergent contains the enzymatic detergent additive according to the invention in an amount of between 0.01 and 50.0% w/w, preferably between 0.1 and 2% w/w. In this manner, a reasonable balance between enzyme action and the action of the other detergent ingredients is generated.

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The third aspect of the invention comprises a washing process in which the detergent used is the detergent according to the invention and in which the pH is between 7 and 11.

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In a specially preferred embodiment of the washing process according to the invention, the washing solution contains the detergent according to the invention in an amount of between 1 and 20 g/l of washing solution. In this manner, a convenient lipase activity is generated in the washing solution, i.e. typically between 1,000 and 5,000 NLU/l of washing solution.

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The invention will be illustrated by the following examples, of which examples 1 -7 are production examples, and examples 8 -13 are washing examples.

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Example 1

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A culture of *Pseudomonas cepacia* DSM 3401 on an agar slant was transferred to five 500 ml shaking flasks, each with 100 ml of Boullion-3 medium, and shaken at 30°C for 1 day (200 rpm, amplitude 2.5 cm).

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The composition of Boullion-3 was as follows:

<u>constituent</u>	<u>concentration, g/l</u>
Peptone	6
Trypsin digested casein	4
Yeast extract	3
Meat extract	1.5
Glucose	1

The medium was autoclaved at 121°C for 40 minutes.

The culture broth of these Boullion-3 shake flasks were used as a seed culture for inoculating two hundred 500 ml shake flasks, each with 200 ml PL-1 medium.

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The composition of the PL-1 medium was as follows:

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<u>constituent</u>	<u>concentration, g/l</u>
Peptone	10
Tween-80	12
MgSO ₄ · 7H ₂ O	2
CaCl ₂ · 2H ₂ O	0.1
pH before autoclaving	6.0

The medium was autoclaved at 121°C. for 40 minutes.

Each PL-1 shake flask was inoculated with 0.5 -2 ml of Boullion-3 culture broth, and shaken with 200 rpm (amplitude 2.5 cm) at 30°C for 5 days. The culture broth from the shake flasks were pooled at harvest, totalling 39.5 l with an enzyme yield of 53 LU/ml.

The culture broth was centrifuged for 35 minutes at 4100 g by means of a Beckman Model J-6 centrifuge. The supernatant was concentrated by ultrafiltration (washed with approximately 1 volume of water) to 1.4 l by a Pellicon ultrafiltration apparatus from Millipore with a 10000 MW cut off filter sheet. The concentrate was freeze dried, and the yield was 56.2 g of powder with a enzyme activity of 21.500 LU/g.

In the following production examples 2 -6 100 ml of medium in a 500 ml shake flask was inoculated with cultures originating from specified strains of *Pseudomonas cepacia* on agar slants incubated at around 30°C for around 3 days. Example 7 is a comparison example representing *Fusarium oxysporum* lipase, obtained as described in example 23 in European patent application with publication No. 0 130 064, and intended for use in example 13, which is a comparative washing example. The medium in examples 2 -6 is LIP7 (described previously).

The shake flasks were incubated for 4 days at around 26°C and at 200 -250 rpm. The lipase activity of the broth (NLU_{8.5}/ml) was measured, and the broth was directly used as a lipase source in a later described washing example.

Production example No.	Strain DSM No.	Lipase activity of		Corresponding washing example No.
		broth NLU _{8.5} /ml	powder NLU _{8.5} /g	
2	3333	13.0		8
3	3334	7.7		9
4	3337	7.1		10
5	3335	10.4		11
6	3336	4.1		12
7	-	-	28300	13

The washing examples 8 -13 were carried out as follows.

Cotton swatches (obtained from the Japan Oil Chemists' Society and designated No. 6) measuring 7.5 x 5.0 cm and weighing 0.52 g were treated with 0.5 ml of 3% beef tallow solution in chloroform on each side.

The swatches were cut into 8 pieces of approximately the same size. These 8 pieces were transferred to a small reaction vessel containing 10 ml of enzyme solution with specified lipase activity (NLU_{8.5}/ml) and with pH 8.5, controlled by means of an approximately 0.2 M buffer of TRIS/HCl (TRIS = 2-amino-2-hydroxymethyl-1,3-propanediol). The reaction mixture was incubated for 1 hour at 40°C with shaking. Then 40 µl of 10% LAS in 0.2 M

borate buffer (pH 10) was added to the reaction mixture. Now the reaction mixture was incubated for 15 minutes at 40°C with shaking. 100 µl of the washing solution was withdrawn for the assay of free fatty acids.

The assay of free fatty acids was carried out colorimetrically in accordance with the method described in the brochure Code 279-75401 Nef C-test Wako obtainable from Wako Chemicals, GmbH, Nissan Strasse 2-4040, Neuussl, West Germany, except that a 100 µl sample was used instead of a 50 µl sample, reagent samples A and B were 0.5 and 1 ml instead of 1 and 2 ml, respectively and that the assay was carried out at 40°C instead of 37°C. A cuvette with a path length

of 1 cm was used for the spectrophotometric measurement. The spectrophotometric reading at 550 nm A, for the washing solution being corrected for the spectrophotometric reading at 550 nm A₀ for a blank (a 100 µl enzyme sample taken out from the reaction vessel before incubation) is the ΔA value ($\Delta A = A_0 - A$) representing the amount of free fatty acids in the washing solution liberated from the swatches. Thus, ΔA is an indication of the lipolytic washing efficiency.

The following washing examples 8 -12 illustrate the invention. Example 13 is the comparative washing example, performed with *Fusarium oxysporum* lipase.

Example No.	Lipase originating from example No.	Lipase acitivity in washing solution, NLU _{8.5} /ml	ΔA
8	2	2.6	0.48
9	3	1.6	0.53
10	4	2.8	0.41
11	5	2.1	0.59
12	6	2.1	0.56
13	7	2.8	0.21

It clearly appears from the above table that the enzymatic detergent additive according to the invention is superior to the *Fusarium oxysporum* lipase, in terms of free fatty acid released from the swatches in the washing solution, at comparable values of lipase activity expressed as NLU_{8.5}/ml.

Deposits DSM 3333 to 3337 and 3401 were all made at the Deutsche Sammlung fur Mikroorganismen under the terms and conditions of the Budapest Treaty. Deposits DSM 3333 to 3336 were made on 28 May 1985, DSM 3337 was made on 10 June 1985 and DSM 3401 was made on 22 July 1985.

The features disclosed in the foregoing description and in the following claims may, both separately and in any combination thereof, be material for realising the invention in diverse forms thereof.

35 Claims

1. Enzymatic detergent additive the active component of which is a microbially produced lipase, wherein the lipase is producible by means of a lipase producing strain of *Pseudomonas cepacia*.
2. Enzymatic detergent additive according to claim 1, wherein the strain is *Pseudomonas cepacia* DSM 3333 to 3337 inclusive, or 3401, preferably DSM 3335 or 3401.
3. Enzymatic detergent additive according to claims 1 to 2, wherein the additive is provided as a non-dusting granulate.
4. Enzymatic detergent additive according to claims 1 to 3, wherein the additive is provided as a liquid.
5. Enzymatic detergent additive according to claim 4, wherein the liquid contains an enzyme stabilizer.
6. Enzymatic detergent additive according to claims 1 to 5, wherein the lipase activity is above about 2500 NLU/g of additive.

7. Enzymatic detergent additive according to claims 1 to 6, wherein the additive besides the lipase contains a proteolytic enzyme.

8. Enzymatic detergent additive according to claim 7, wherein the proteolytic activity is between about 0.5 and about 3.0 Anson units/g of additive.

9. Detergent comprising an enzymatic additive, the active component of which is a microbially produced lipase, wherein the enzymatic detergent additive is the enzymatic additive according to claims 1 to 8.

10. Detergent according to claim 9, wherein the detergent contains the enzymatic detergent additive according to claim 1 to 10 in an amount of between 0.01 and 50.0% w/w, preferably between 0.1 and 2% w/w.

5 11. Washing process, wherein the detergent is the detergent according to claims 9 -10 and the pH is between 7 and 11.

10 12. Washing process according to claim 11, wherein the washing solution contains the detergent according to claims 11 to 12 in an amount of between 1 and 20 g per litre of washing solution.

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